



Agriculture and
Agri-Food Canada

Agriculture et
Agroalimentaire Canada



Aerobiological Surveillance of Wheat Pathogens

Laroche, A.¹, G.T. Araujo^{1, 2}, W. Lu³, E. Amundsen¹, N.K. Newlands³, R. Aboukhaddour¹, J. Larsen¹, H. Rhandawa¹, R. Graf¹, D.A. Gaudet¹, B.L. Selinger², M. Frick¹

¹Agriculture and Agri-Food Canada, Lethbridge RDC

²University of Lethbridge, Department of Biological Sciences, Lethbridge

³Agriculture and Agri-Food Canada, Summerland RDC

Red Deer, Alberta January 10, 2018

Canada 

Presentation Outline

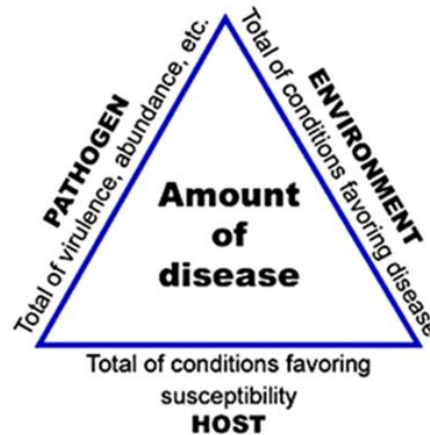
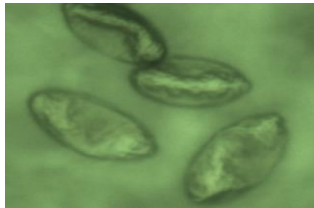
- Wheat pathogen surveillance tools
- Collection of samples
- Detection of samples
- Disease Modeling
- Future prospects



Puccinia striiformis f.sp. tritici (Pst) causal agent of stripe (yellow) rust

Introduction

- Host-pathogen interaction
 - Not all interactions are successful
 - Pathogens can cause mild to severe damage
 - Disease Triangle Concept



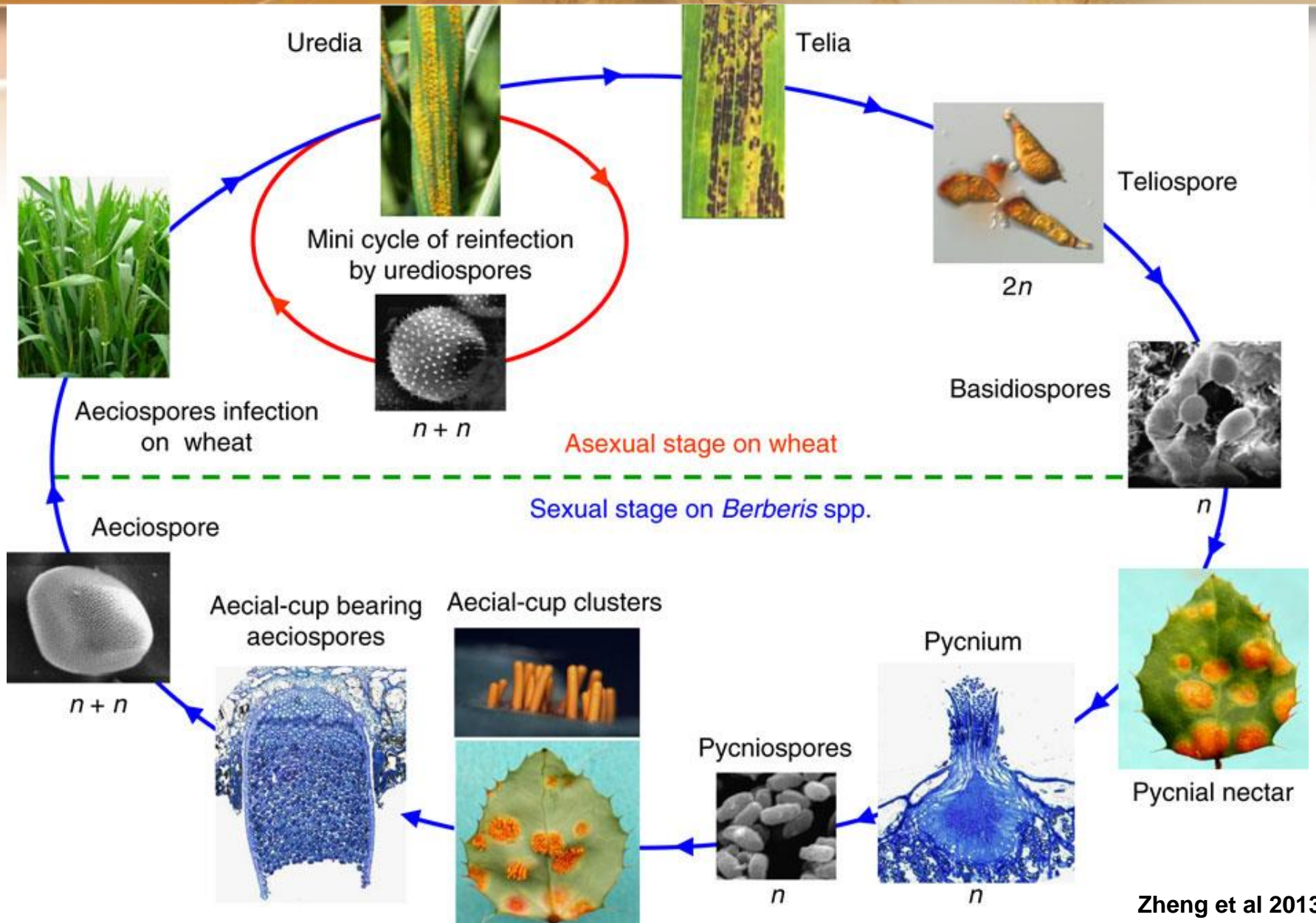
Credits: <http://www.outdoorphotographycanada.com>

Credits: <https://masters.agron.iastate.edu>



Credits: <http://www.watermarkchurch.hk>

Puccinia striiformis f.sp. *tritici* life cycle



Wheat pathogen surveillance tools

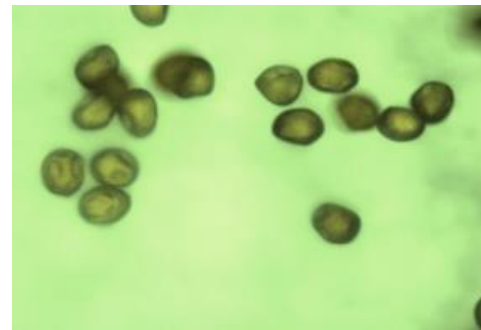
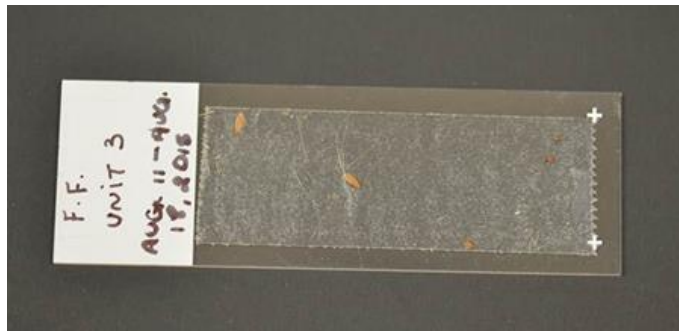
- **Currently, we have to wait for symptoms on susceptible lines of wheat**



Wheat pathogen surveillance tools

- We are using sticky slides to detect presence of potential airborne pathogens.

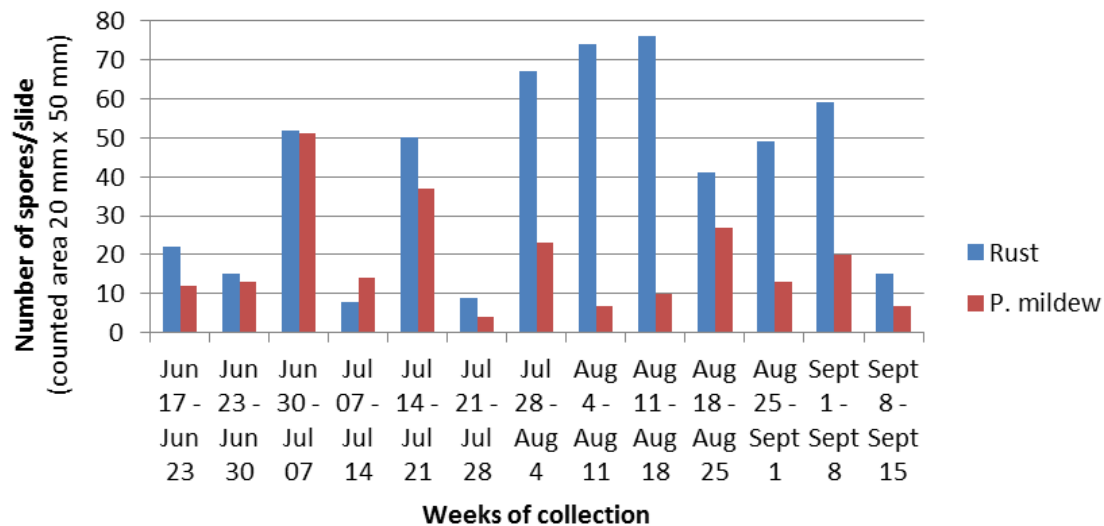
Adhesive tape



rust spores – 400X

Microscopy analyses

Spores Counting - Weather Station 2016



Wheat pathogen surveillance tools

- We are also using spore collectors to detect presence of potential airborne pathogens.

Burkard Cyclone



Location of spore collector units in southern Alberta



* 2016 & 2017

¥ 2016

⌘ 2017

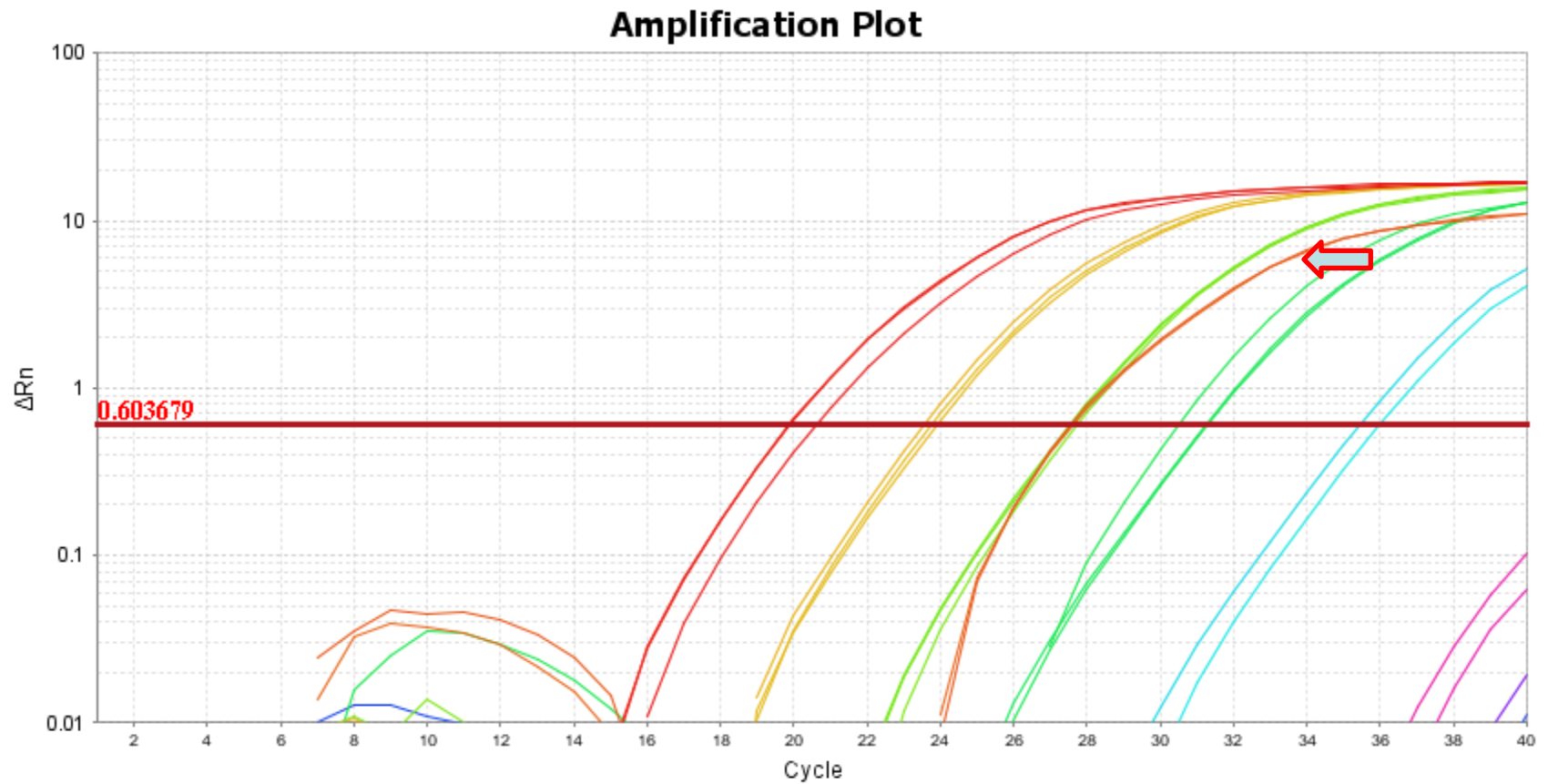
Detection of samples

- **DNA level**
 - Different standard approaches enable identification at species level, very sensitive and highly specific
- **Immunological level**
 - Identification at species level, very sensitive, highly specific and rapid

Detection of samples

- **DNA level**
 - Need to break the spore to release the DNA and then detect the presence of a given pathogen based on a unique DNA sequence.
 - DNeasy PowerSoil DNA Isolation Kit (Qiagen) a 2 step protocol that first breaks the spores open and then yield a fraction of DNA that can be used to detect pathogens.
 - Identified and validated unique DNA sequences for each species
 - qPCR to detect and quantify pathogens.

qPCR results (stripe rust target)



Pathogen-specific primer sets

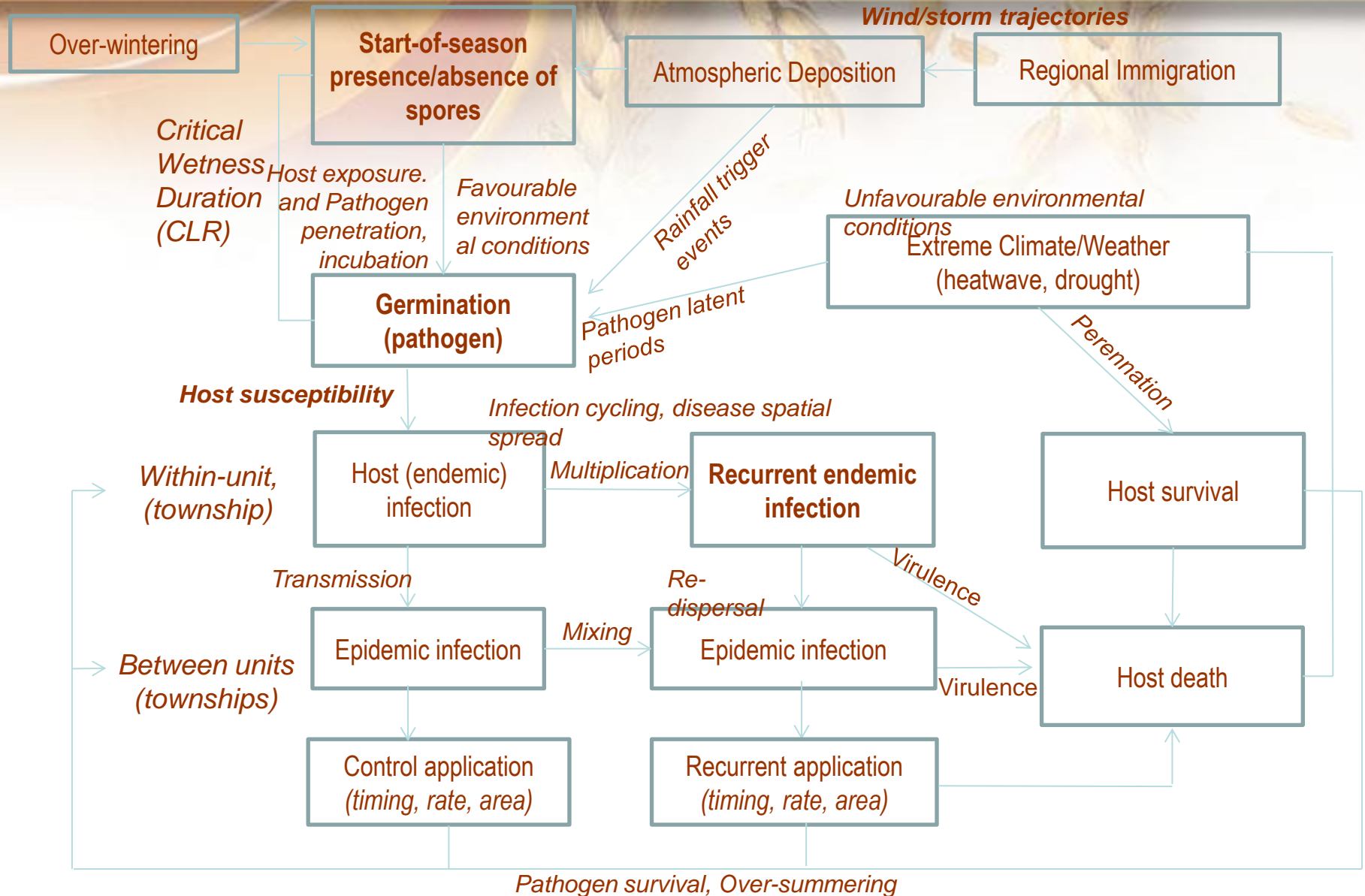
- The seven best PCR primer sets tested

Primer set (F/R)	Pathogens	Diseases	Detection level
Pg2	<i>Puccinia graminis f. sp. tritici</i>	Stem rust	0.2 pg - 16 spores
PST	<i>Puccinia striiformis f. sp. tritici</i>	Stripe rust	0.3 pg - 25 spores
Pt2-032/ Pt1-113#2	<i>Puccinia triticina f. sp. tritici</i>	Leaf rust	0.5 pg - 20 spores
Bgt -6	<i>Blumeria graminis f. sp. tritici</i>	Powdery mildew	2.0 pg - 307 spores
Tox A1/ Tox A2	<i>Pyrenophora tritici-repentis</i>	Tan spot	0.1 pg - 7 spores
Fgram B	<i>Fusarium graminearum</i>	Fusarium head blight	0.2 pg - 8400 spores
ITS1/ITS4	Universal primer	All fungal samples	-----

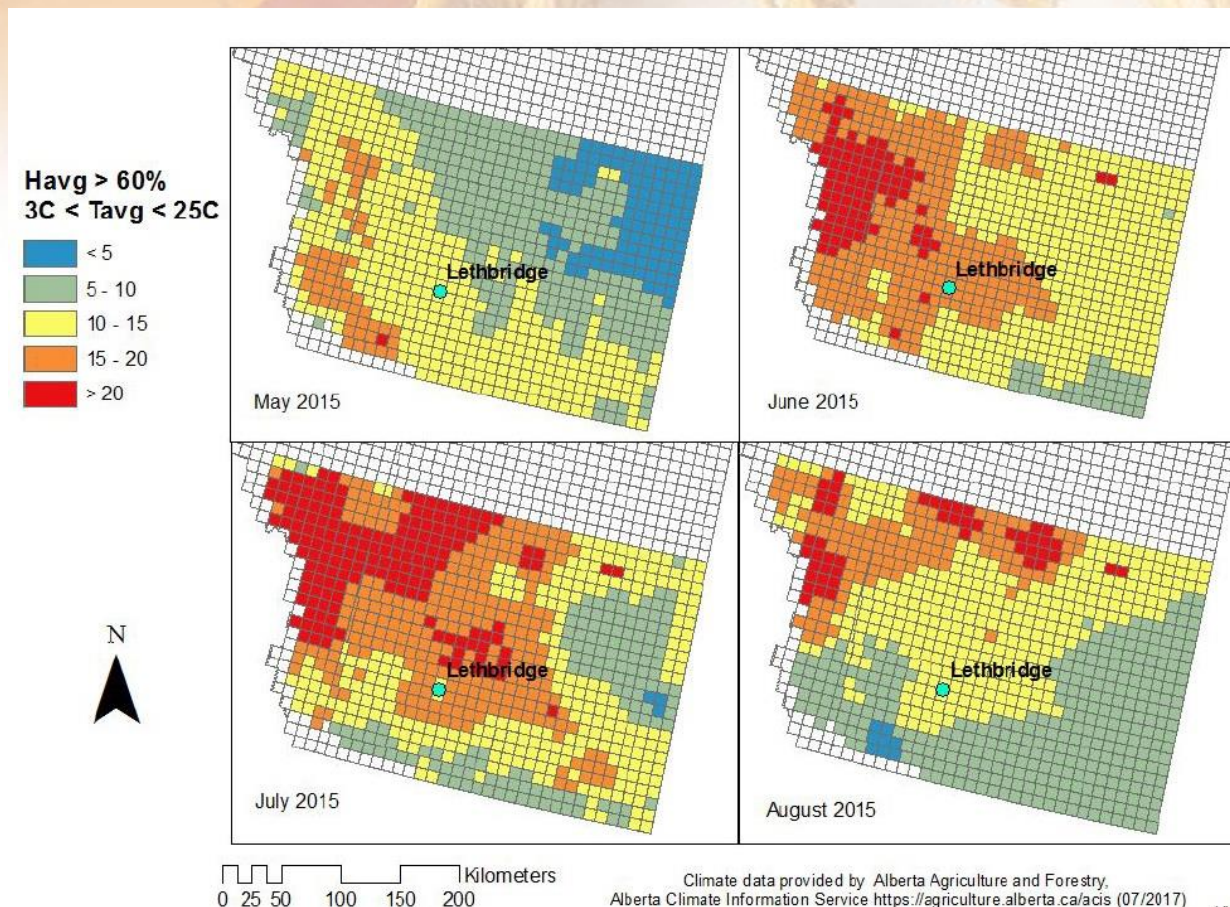
Summary

- We can reliably detect 6 wheat pathogens at a very low detection level: stripe rust, stem rust, leaf rust, powdery mildew, tan spot and Fusarium head blight
- The assay is rapid and would covers the most prevalent diseases of wheat on the Canadian Prairies
- Detection level well below minimum number of spores needed for successful infection at field level.

Disease modeling and forecasting framework



Potential wheat rust disease occurrence based on temperature and humidity empirical thresholds (Alberta municipalities, 2015)



***municipality-interpolated high-resolution climate data
from Alberta Climate Information Service (ACIS), <https://agriculture.alberta.ca/acis/>**

Spatial endemic epidemic multivariate time-series model (HHH4)

Assumptions:

- Negative binomial model
- Constant population
- Districts have the same seasonality effect at time t
- Autoregressive and epidemic effects are homogeneous across districts, constant in time
- Spores only spread from the side neighboured districts

Basic model (without covariate spatial random effects):

$$\mu_{it} = \underbrace{e_i \nu_t}_{\text{endemic}} + \underbrace{\lambda Y_{i,t-1}}_{\text{autoregressive}} + \underbrace{\phi \sum_{j \neq i} w_{ji} Y_{j,t-1}}_{\text{epidemic}}$$

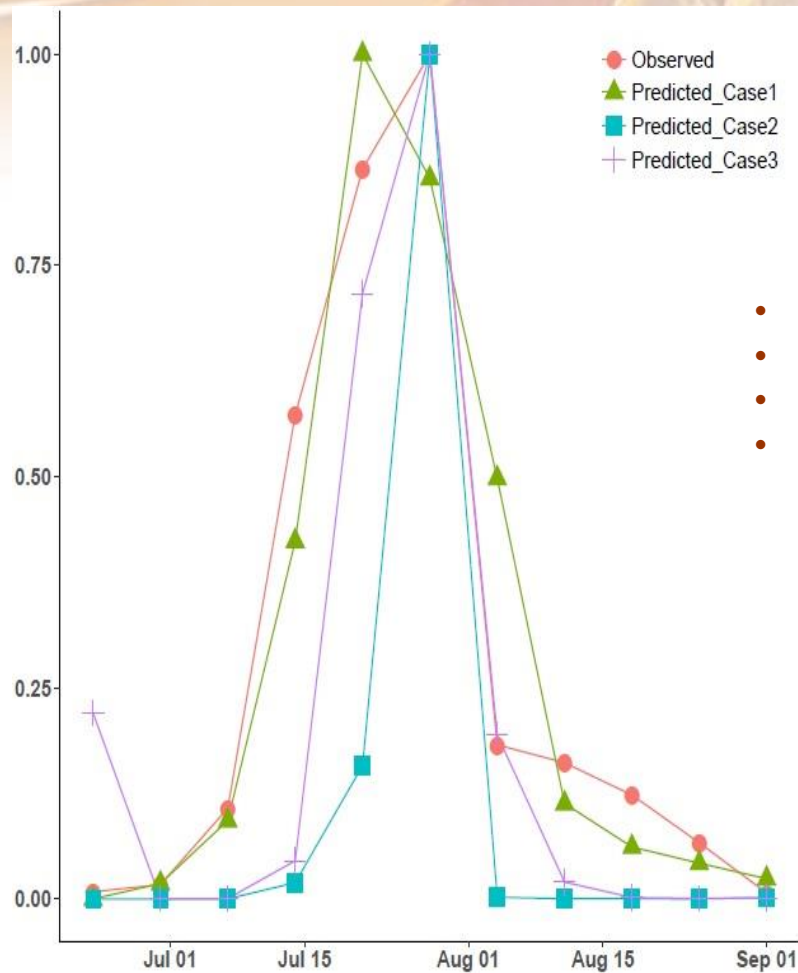
$$\log(\nu_t) = \underbrace{\alpha^{(\nu)}}_{\text{initial states}} + \underbrace{\beta_t t}_{\text{gradient}} + \underbrace{\gamma \sin(\omega t) + \delta \cos(\omega t)}_{\text{seasonal trend}}$$

Where

- e_i is the population for district i .
- ν_t is the seasonality effect.
- α and $\exp(\beta_t)$ are the **endemic** parameters.
- λ is the **autoregressive** parameter, $\lambda = \exp(\alpha^{(\lambda)})$.
- ϕ is the **epidemic** parameter, $\phi = \exp(\alpha^{(\phi)})$.

Spatial (HHH4) model predictions

Infection count (unitless)



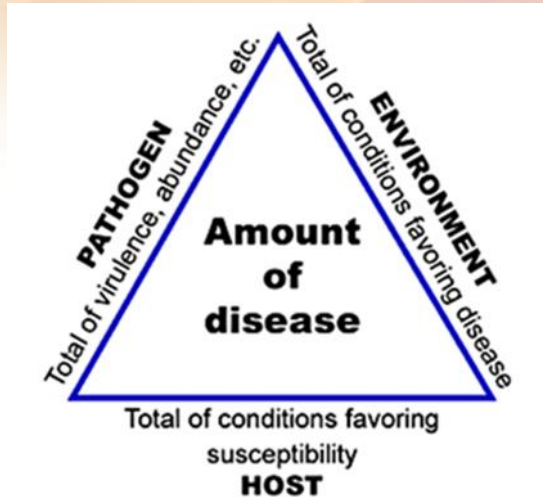
Time (weeks)

- Observed spore counts (Lethbridge, 2015)
- Case 1 (HHH4 model, observed climate data)
- Case 2 (HHH4 model, ensemble-based reanalysis data)
- Case 3 (HHH4 model, observed and reanalysis data)

Summary

- **Spatial model is able to predict disease occurrence width and peak timing well (Fairfield, Lethbridge validation site)**
- **Highest model prediction is achieved for the spatial model (HHH4) that assumes disease spread between adjacent districts**
- **Prediction power increases when combining station measured and ensemble-model reanalysis climate information**
- **Forecast model development/validation requires larger sample size of disease monitoring data, multiple growing seasons (i.e., larger network of cyclone air samplers needed)**

Future Prospects



+

**Modelization
of infection
of stripe rust
and other
diseases**



Near real-time information to help in decision making toward mitigating impact of pathogens on wheat yield and quality

Credits: <https://masters.agron.iastate.edu>

Acknowledgements

The background of the slide features a soft-focus image of several golden wheat stalks and some loose grains. In the upper left corner, a portion of a bowl with a yellow and pink rim is visible. The overall color palette is warm, dominated by yellows, oranges, and soft pinks.

AAFC Peer Reviewed Projects & GRDI

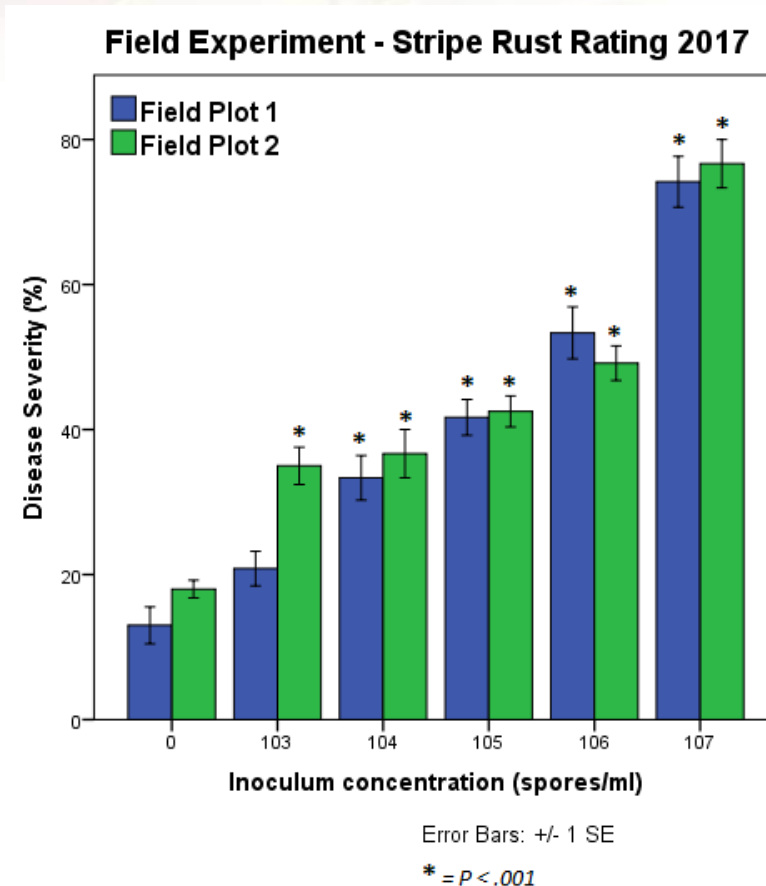


?



Previous experiment

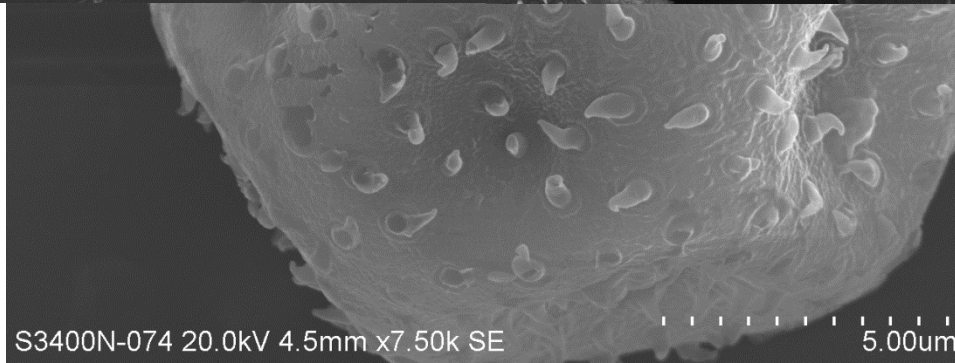
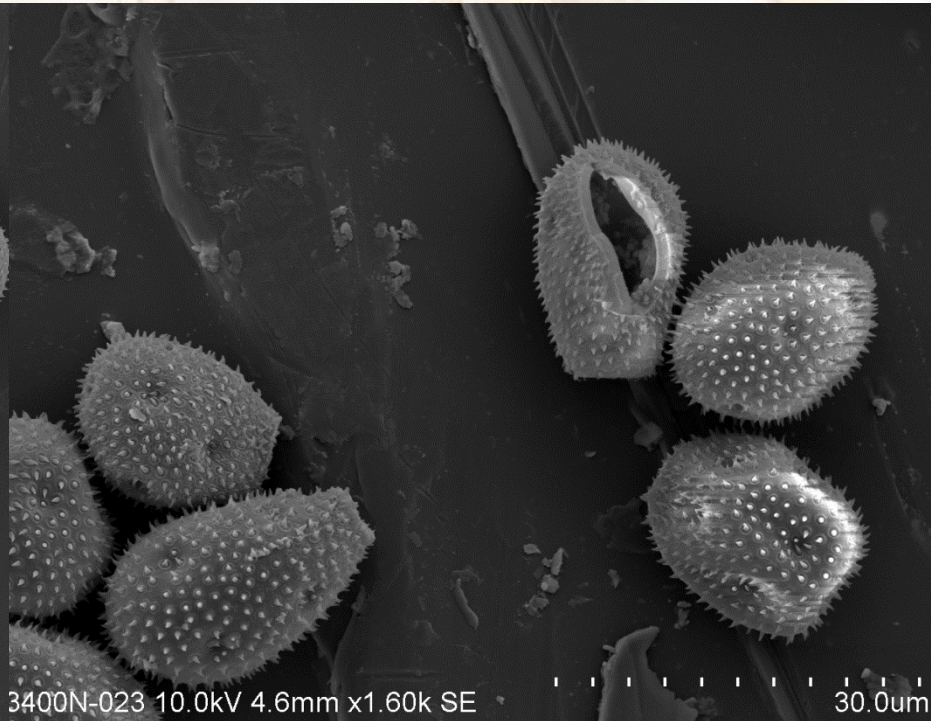
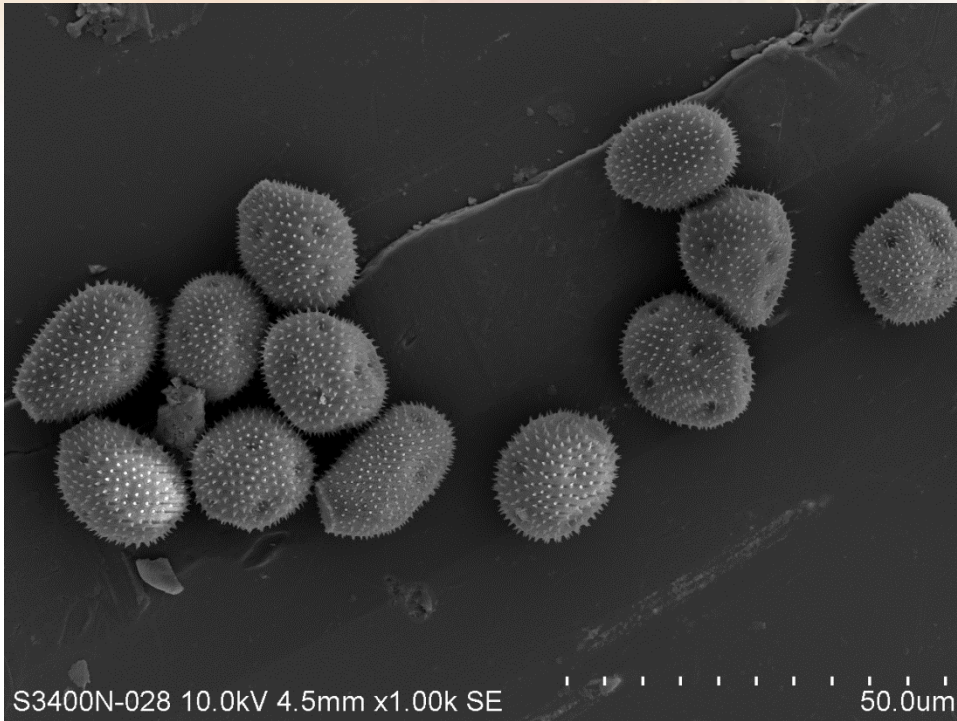
- Field experiment – stripe rust



Detection of samples

- **Light Microscopy**
 - Limited resolution and best approximation at genus level
- **Electron Microscopy**
 - Additional information but slow process

Pst spores under scanning electron microscope



Courtesy C. Sheedy LRDC

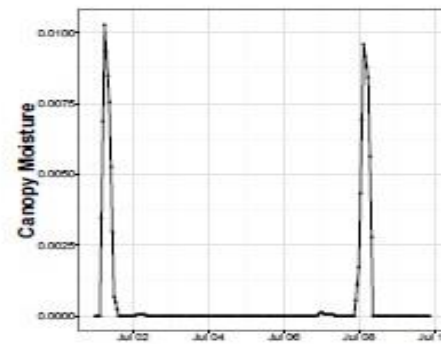
Wheat stripe rust - Modeled germination and infection rate

(Lethbridge, 2015 growing season)

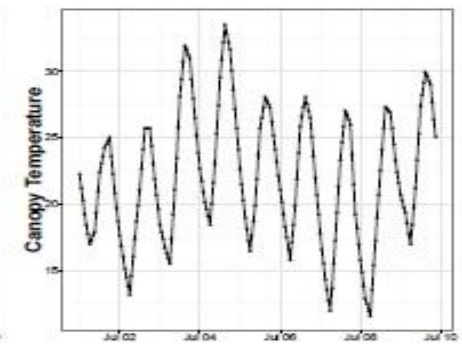
Infection profile (site-specific distribution) (CLR model)

Assumptions:

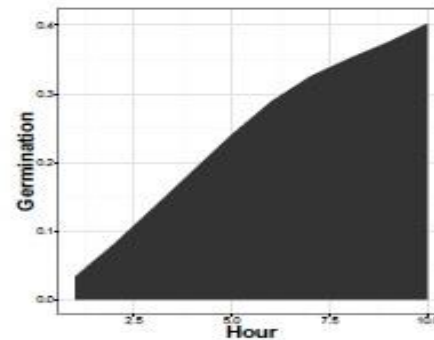
- Temperature is constant over each hourly interval
- Equal spore cohorts germinating at the start of each wet hour
- No spore germination process during the dry periods
- No neighbouring infections
- Spore germination, infection are only affected by canopy temperature and moisture
- The process of spore germination and infection is Weibull-distributed



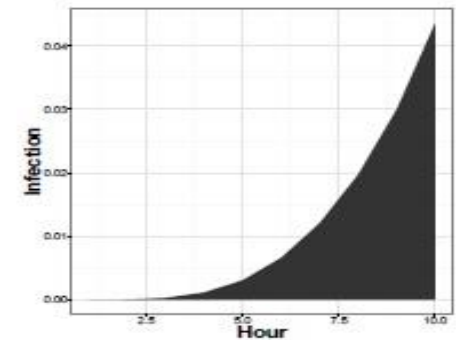
(a) Moisture



(b) Temperature



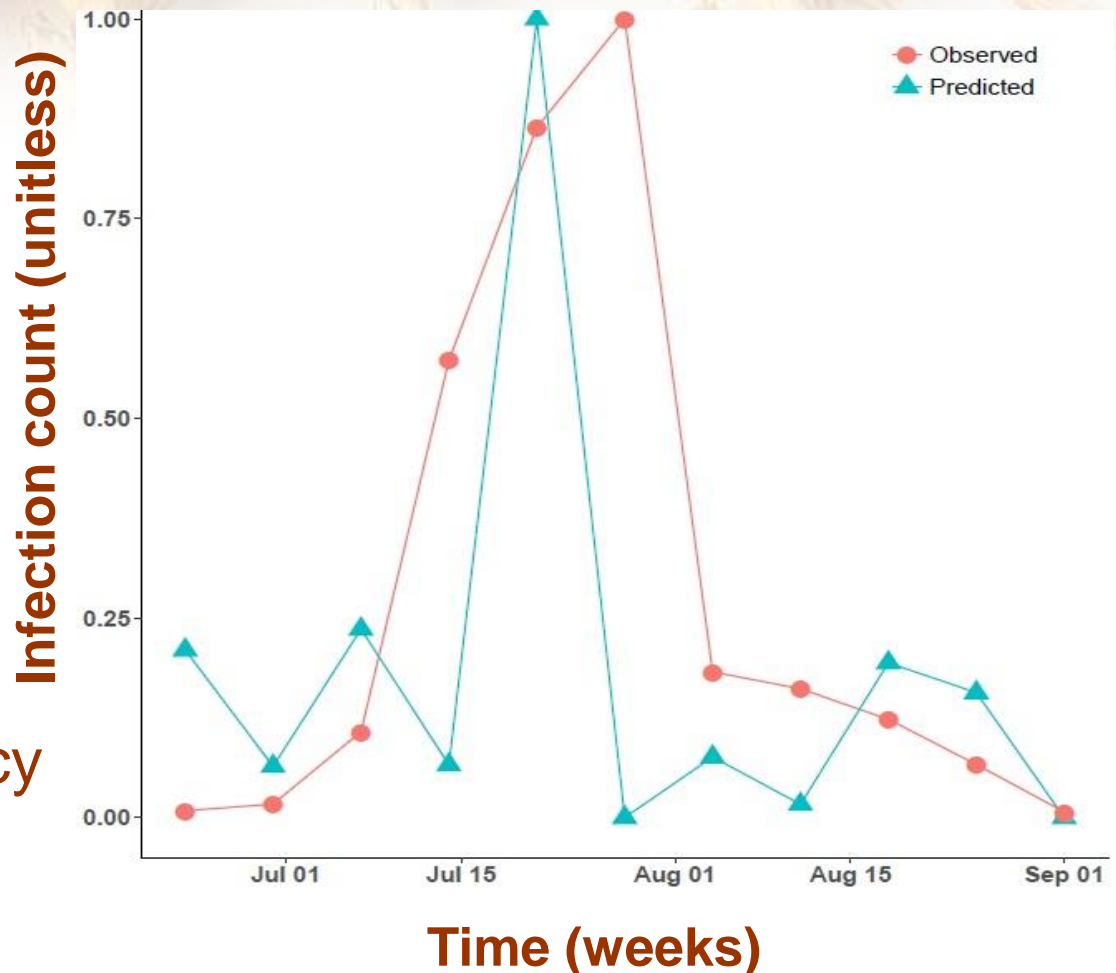
(c) Germination rate



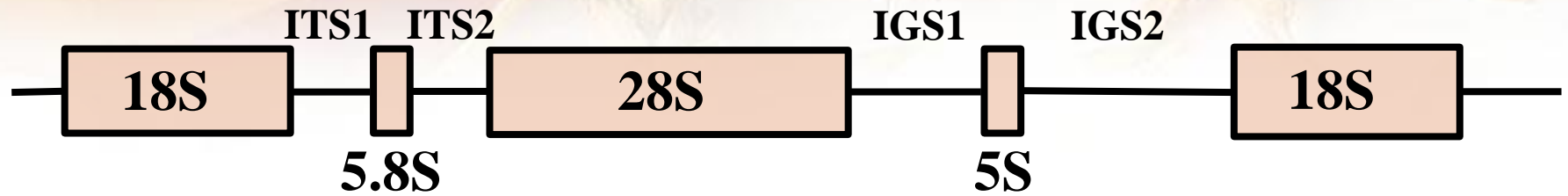
(d) Infection rate

CLR model predictions (normalized) (2015 growing season, Lethbridge spore profile)

- higher variability in prediction error during early/late season
- observed infection rate close to predicted
- timing of infection shows largest discrepancy



Schematic of rDNA locus



Fungi placement

Eukaryota

Kingdom: Fungi

Phylum: Basidiomycota

Class: Agaricomycetes

Subclass: Agaricomycetidae

Order: Atheliales

Family: Atheliaceae

Genus: *Athelia*

Species: *A. arachnoidea*

Fungi

Ascomycota

Sordariomycetes

Xylariomycetidae

Xylariales

Microdochiaceae

Microdochium

M. nivale

Wheat rusts

stripe rust



stem rust



leaf rust

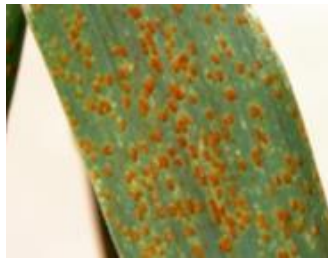


Common diseases of wheat in Western Canada

- Stripe rust (*Puccinia striiformis* f. sp. *tritici*)
- Leaf rust (*Puccinia triticina*)
- Stem rust (*Puccinia graminis* f. sp. *tritici*)
- Powdery mildew (*Blumeria graminis* f. sp. *tritici*)
- Tan spot (*Pyrenophora tritici-repentis*)
- Fusarium head blight (*Fusarium graminearum*)



<http://msue.anr.msu.edu>



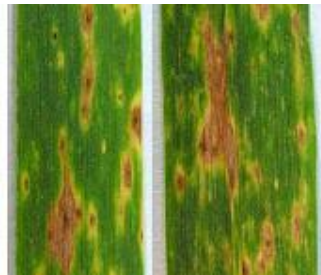
<http://theconversation.com>



<http://agric.wa.gov.au>



<http://agric.wa.gov.au>



<http://www.grdc.com.au>



<https://www.ag.ndsu.edu>

Wheat rusts

- Rusts are obligate biotrophs
- *Puccinia striiformis f.sp. tritici* (*Pst*) is causal agent of stripe rust 117 Mb
- *Puccinia triticina* (*Pt*) is causal agent of leaf rust 135 Mb
- *Puccinia graminis f.sp. tritici* (*Pgt*) is causal agent of stem rust 89 Mb
- Around the world, wheat rusts can cause yield reduction from 20% to 90%

Genome size of different organisms and number of genes

	Size (Mb)	Genes (1000)
<i>Escherichia coli</i> (bacteria)	4.7	3
<i>Saccharomyces cerevisiae</i> (yeast)	15	6
<i>Tilletia caries</i> (bunt)	23	
<i>Arabidopsis thaliana</i> (mouse cress)	70	23
<i>Puccinia striiformis</i> (stripe rust)	117	
<i>Drosophila melanogaster</i> (fruit fly)	140	15
<i>Oryza sativa</i> (rice)	580	27
<i>Brassica napus</i> (rapeseed)	1,200	
<i>Medicago sativa</i> (alfalfa)	1,600	
<i>Zea mays</i> (maize)	2,400	
<i>Homo sapiens</i> (man)	3,300	30
<i>Hordeum vulgare</i> (barley)	5,100	
<i>Thinopyrum elongatum</i> (2x)	6,000-8,000?	
<i>Secale cereale</i> (rye)	8,300	
<i>Avena sativa</i> (oat)	11,300	
<i>Triticum aestivum</i> (wheat)	16,000	57
<i>Fritillaria assyriaca</i> (lily)	123,000	

Arumuganathan and Earle, 1991; Ausubel et al. 1995; Brown 1991;
Dean and Schmidt, 1995; Mills 1996.

Acknowledgements

Associates and collaborators at LeRDC:

**Michele Frick
Alejandro Peña
Briana Smith
Eric Amundsen**

**Denis Gaudet
Harpinder Randhawa
Robert Graf
Jamie Larsen**

**Ottawa RDC:
Scott Redhead
Hélène Labbé**

AAFC Peer Reviewed Projects

Alberta Crop Industry Development Fund